

Fig. 1 The observed pseudo-first-order rate constant for the alcoholysis of cephaloridine in water at 30 °C, $I = 1.0$ M (KCl) as a function of total hexafluoropropan-2-ol concentration at the indicated pH

group at C-7, *i.e.* 1: X = NH₂, with cephalosporins lacking a leaving group at C-3 *i.e.* 1: L = H, with cephalosporins having an ester function at C-4 rather than a carboxylate and also with Δ^2 -cephalosporins. It is, therefore, concluded that the observation is an intrinsic property of the alcoholysis reaction.

In the lower pH range of the buffer system the concentration of undissociated alcohol increases and it was assumed that there may be a general acid-catalysed inhibition of the reaction. This was confirmed by adding general acids such as higher pK_a alcohols or ammonium salts to the alcohol buffer system. At the higher pH range where 'normal' behaviour is observed adding these general acids causes the onset of inhibition, whereas in the lower pH region, in which the slower rate is observed, adding these additional proton donors has no effect (Fig. 3).

We can think of no explanation other than that the anionic tetrahedral intermediate T⁻ formed by nucleophilic attack of the alkoxide ion on the β -lactam is trapped by proton donors to form the thermodynamically more stable but less reactive neutral intermediate T⁰ (Scheme 1). Even at pH below the pK_a of T⁰ the intermediate T⁻ cannot be protonated if the rate constant for this step is lower than the breakdown of T⁻ to reactants and products. The half-life of anionic tetrahedral intermediates⁶ is commonly of the order of 10⁻⁸ to 10⁻¹¹ s and the longer the lifetime the more likely is the intermediate to be protonated by acids at the diffusion controlled limit, $k_3[\text{HA}]$ (Scheme 1). Tetrahedral intermediates of carbonyl carbon in four-membered rings are expected to be relatively more stable than their acyclic analogues.^{1,7} The opening of four-membered rings is not the facile process expected from the release of strain energy.^{1,8} It is, therefore, not surprising that the tetrahedral intermediate, T⁻, formed from the alkoxide-

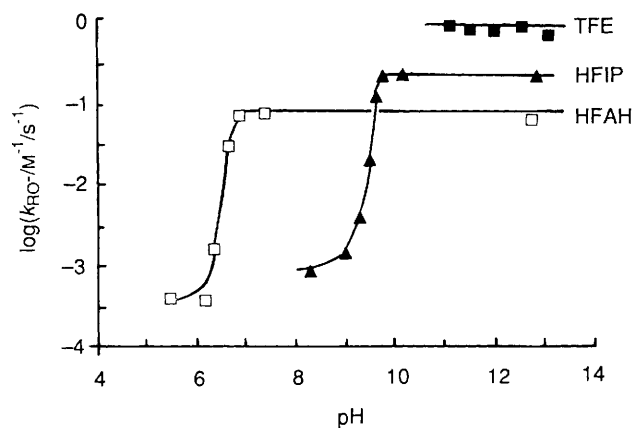


Fig. 2 The second-order rate constant for alkoxide ions reacting with cephaloridine in water at 30 °C $I = 1.0$ M (KCl) as a function of pH; TFE = trifluoroethanol, HFIP = hexafluoropropan-2-ol and HFAH = hexafluoroacetone hydrate

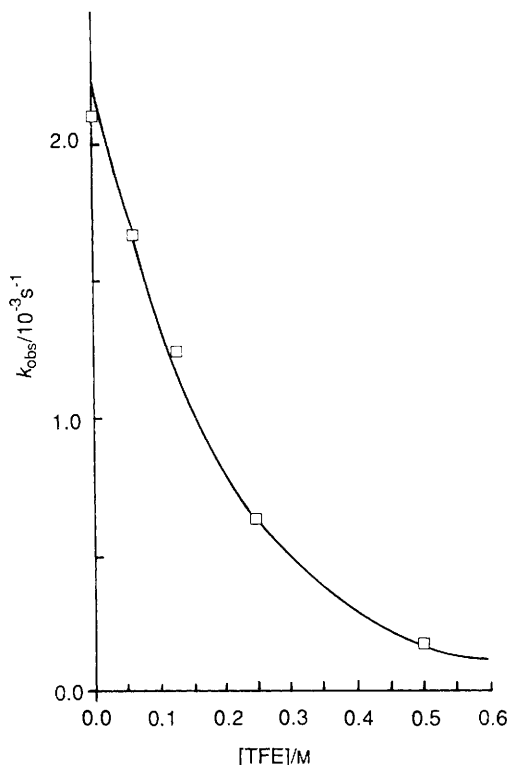


Fig. 3 The effect of increasing trifluoroethanol concentration on the observed pseudo-first-order rate constant for the alcoholysis of the 3-methyl cephalosporin (1, L = CH₃) in 0.2 M hexafluoropropan-2-ol buffer pH 10.25 at 30 °C, $I = 1.0$ M (KCl)

ion attack on cephalosporins has a sufficiently large lifetime to allow protonation to occur.

In the high pH region of the buffers the rate of breakdown of T⁻ is faster than protonation, $(k_{-1} + k_2) > k_3[\text{HA}]$. Under these conditions the second-order rate constants for alkoxide-ion attack, k_{RO^-} , generate Brønsted β_{nuc} values of 0.2 to 0.3; consistent with rate-limiting formation of the tetrahedral intermediate. At lower pH, or in the presence of additional proton donors, protonation of T⁻ to T⁰ becomes competitive with breakdown of T⁻ and $k_3[\text{HA}] > (k_{-1} + k_2)$. Under these conditions the Brønsted β_{nuc} values are 0.8 to 1.0 and are

consistent with the rate-limiting breakdown of the intermediate.

There is some 270 MHz NMR evidence that the tetrahedral intermediate actually accumulates. In trifluoroethanol buffers at low pH, but not at high pH, the 3-methyl cephalosporin, **1** (L = H, X = PhCH₂CONH), shows high field shifts of the β -lactam 6-H and 7-H from an AB 'quartet' centred at δ 5.06 and 5.56 ppm with $J_{6,7}$ 4.3 Hz to two different AB 'quartets' with different intensities centred at δ 4.52 and 4.62 ($J_{6,7}$ 9.85 Hz) and 4.55 and 4.65 δ ($J_{6,7}$ 7.88 Hz). There are two new methyl resonances at δ 1.88 and 1.92 and the H-2 protons also move upfield from δ 3.24 and 3.61, $J_{\alpha,\beta}$ 17.2 Hz, to a doublet at δ 3.10 but a pair of doublets at δ 3.38 and 3.45, $J_{\alpha,\beta}$ 16.8 Hz. These spectral changes are consistent with the formation of the two stereoisomers of the tetrahedral intermediate T⁰ formed in a ratio of ca. 60:40. Only β protons of H-2 are affected by the stereochemistry at the tetrahedral intermediate.

It has been previously suggested that although electrophilic catalysis at the carbonyl oxygen may enhance nucleophilic attack at carbon, complete neutralisation of the incipient charge on oxygen is inhibitory for expulsion of the leaving group from the carbonyl centre.⁹ This is illustrated by the present example where the decomposition of the neutral tetrahedral intermediate T⁰ is several hundredfold slower than T⁻.

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References

- 1 M. I. Page, *Adv. Phys. Org. Chem.*, 1987, **23**, 165; J. M. Frere and B. Joris, *CRC Crit. Rev. Microbiol.*, 1985, **11**, 299; W. Durckheimer, J. Blumbach, R. Lattrell and K. H. Scheunemann, *Angew. Chem., Int. Ed. Engl.*, 1985, **24**, 180.
- 2 M. L. Bender, *Chem. Rev.*, 1960, **60**, 53; S. L. Johnson, *Adv. Phys. Org. Chem.*, 1967, **5**, 237; W. P. Jencks, *Catalysis in Chemistry and Enzymology*, McGraw-Hill, New York, 1969, pp. 473-554; A. Williams, *Acc. Chem. Res.*, 1989, **22**, 387.
- 3 M. L. Bender, *J. Am. Chem. Soc.*, 1951, **73**, 1626.
- 4 B. Capon, A. K. Ghosh and D-M. A. Grieve, *Acc. Chem. Res.*, 1981, **14**, 306; B. Capon, M. I. Dosunnu and M. de N. Matos-Sanchez, *Adv. Phys. Org. Chem.*, 1985, **21**, 37; R. A. McClelland and L. J. Santry, *Acc. Chem. Res.*, 1983, **16**, 394.
- 5 W. P. Jencks, *Acc. Chem. Res.*, 1976, **9**, 425; *Acc. Chem. Res.*, 1980, **13**, 161.
- 6 N. P. Gensmantel and M. I. Page, *J. Chem. Soc., Perkin Trans. 2*, 1979, 137; M. I. Page and P. Proctor, *J. Am. Chem. Soc.*, 1984, **106**, 3820; M. I. Page, and W. P. Jencks, *J. Am. Chem. Soc.*, 1972, **94**, 8828.
- 7 M. I. Page, *Chem. Soc. Rev.*, 1973, **2**, 295.
- 8 M. I. Page, P. Webster and L. Ghosez, *J. Chem. Soc., Perkin Trans. 2*, 1990, **805**, 813; M. I. Page, P. Webster, L. Ghosez and S. Bogdan, *Bull. Soc. Chim. Fr.*, 1988, 272; M. A. Cadadei, A. di Martino, C. Galli and L. Mandolini, *Gazz. Chim. Ital.*, 1986, **116**, 659; D. F. De Tar and W. Brooks, *J. Org. Chem.*, 1978, **43**, 2245; H. A. Earl and C. J. M. Stirling, *J. Chem. Soc., Perkin Trans. 2*, 1987, 1273.
- 9 N. P. Gensmantel, P. Proctor and M. I. Page, *J. Chem. Soc., Perkin Trans. 2*, 1980, 1725; M. I. Page, *The Chemistry of Enzyme Action*, ed., M. I. Page, Elsevier, Amsterdam, 1984, 229.